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| --- | --- |
| **Hashcode:** | |
| Collected By: | Date of Collection: |
| Time specimen collected: | |
| Location of Box -80: | |

|  |  |
| --- | --- |
| **Patient Information/Sticker:** | |
| Name: | DOB |
| MRN: | Sex: |
| Race: | |
| **REDCap:** | **OnCore:** |

* Bleach beaker
* 5 x 5 mL glass tubes
* 8 x 2 mL plastic tubes
* Incubation Period: >30 minutes, but <40 minutes
* Total Processing Time: Under 3 hours (from blood draw to freezer)
  + Biopsy: Yes / No
  + Genetic Patient: Yes / No / TBD

**Tube # 2 (8.5 mL Red/Black Top Tube, Serum Separator)**

1. Invert very slowly 6-8 times to mix, then balance centrifuge exactly with counterweight tube if needed. (Counterweight should be of the same type of tube but filled with water)
2. Centrifuge at 1,000 G for 15 minutes at 23 C. Record start time:\_\_\_\_\_\_\_\_\_

3. Transfer clear supernatant into n=1 glass tube (5 mL capacity)

**Don’t aspirate buffy coat**

-if buffy coat is aspirated, re-spin at previous instructions, and document as ‘re-spun’

4. Aliquot as follows:

* 1. 200 uL x 6 tubes
  2. 500 uL x 5 tubes

5. Cap the cryotubes.

6. Wipe with paper towel, store at -80 °C. Record end time:\_\_\_\_\_\_

7. Dispose original blood tube in a biohazard sharps container.

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| --- | --- | --- |
| **Amount aliquoted** | **Initial Tube** | **Final Tube** |
| **200 uL** |  |  |
| **500 uL** |  |  |

**Tube # 1 (10 mL Purple Top, EDTA)**

1. Invert very slowly 6-8 times to mix, then balance centrifuge exactly with counterweight tube if needed. (Counterweight should be of the same type of tube but filled with water)
2. Centrifuge at 1,000 G for 15 minutes at 23 C. Record start time:\_\_\_\_\_\_\_\_\_
3. Transfer clear plasma into two 2 mL capacity microcentrifuge tubes
4. **Don’t aspirate buffy coat**

-if buffy coat is aspirated, re-spin at previous instructions, and document as ‘re-spun’

1. Centrifuge at 10,000 G for 10 minutes at 4 C.
2. Combine supernatant from both microcentrifuge tubes into n=1 glass tube (5 mL capacity)
3. Aliquot as follows:
   1. 400 uL x 1 tube
   2. 200 uL x 15 tubes
4. Cap the cryotubes.
5. Wipe with paper towel, store at -80 °C. Record end time:\_\_\_\_\_\_
6. Dispose original blood tube in a biohazard sharps container.

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| **Amount aliquoted** | **Tube Number** |
| **400 uL** |  |

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| **Amount aliquoted** | **Initial Tube** | **Final Tube** |
| **200 uL** |  |  |

**Tube # 3 (6 mL Yellow Top Tube, ACD)**

*For Biobanking*:

1. With remainder of blood, invert very slowly 6-8 times to mix, then balance centrifuge exactly with counterweight tube if needed.
2. Centrifuge at 1,000 G for 15 minutes at 23 C. Record start time:\_\_\_\_\_\_\_\_\_
3. Transfer clear supernatant into two 2 mL capacity microcentrifuge tubes
4. Centrifuge at 10,000 G for 10 minutes at 4 C.
5. Combine supernatant from both microcentrifuge tubes into n=1 glass tube (5 mL capacity)
6. Aliquot supernatant as follows:
   1. 200 uL x 6 tubes
   2. 500 uL x 5 tubes
7. Cap the cryotubes.
8. Wipe with paper towel, store at -80 °C. Record end time:\_\_\_\_\_\_
9. Dispose original blood tube in a biohazard sharps container.

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| **Amount aliquoted** | **Initial Tube** | **Final Tube** |
| **200 uL** |  |  |
| **500 uL** |  |  |

**Tube # 4 (4 mL Green Top Tube, Lithium Heparin)**

1. Invert very slowly 6-8 times to mix, then balance centrifuge exactly with counterweight tube if needed. (Counterweight should be of the same type of tube but filled with water)
2. Centrifuge at 1,000 G for 15 minutes at 23 C. Record start time:\_\_\_\_\_\_\_\_\_
3. Transfer clear supernatant into two 2 mL capacity microcentrifuge tubes
4. **Don’t aspirate buffy coat**

-if buffy coat is aspirated, re-spin at previous instructions, and document as ‘re-spun’

1. Centrifuge at 10,000 G for 10 minutes at 4 C.
2. Combine supernatant from both microcentrifuge tubes into n=1 glass tube (5 mL capacity)
3. Aliquot as follows:
   1. 200 uL x 6 tubes
   2. 500 uL x 2 tubes
4. Cap the cryotubes.
5. Wipe with paper towel, store at -80 °C. Record end time:\_\_\_\_\_\_
6. Dispose original blood tube in a biohazard sharps container.

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| **Amount aliquoted** | **Initial Tube** | **Final Tube** |
| **200 uL** |  |  |
| **500 uL** |  |  |

**Tube # 5 (Two 2.7 mL Blue Top Tubes, Sodium Citrate)**

1. Centrifuge at 1,000 G for 15 minutes at 23 C. Record start time:\_\_\_\_\_\_\_\_\_
2. Transfer clear plasma into two 2 mL capacity microcentrifuge tubes
3. **Don’t aspirate buffy coat**

-if buffy coat is aspirated, re-spin at previous instructions, and document as ‘re-spun’

1. Centrifuge at 10,000 G for 10 minutes at 4 C.
2. Combine supernatant from both microcentrifuge tubes into n=1 glass tube (5 mL capacity)
3. Aliquot as follows:
   1. 200 uL x 6 tubes
   2. 500 uL x 3 tubes
4. Cap the cryotubes.
5. Wipe with paper towel, store at -80 °C. Record end time:\_\_\_\_\_\_

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| --- | --- | --- |
| **Amount aliquoted** | **Initial Tube** | **Final Tube** |
| **200 uL** |  |  |
| **500 uL** |  |  |